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Practical Construction of Taxoid C-Ring Segments with a Consecutive Heterodomino Transformation as the Key Reaction Step

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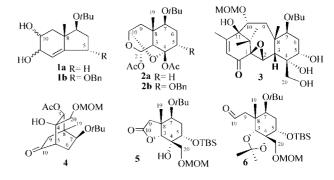
An efficient preparation of fully functional, optically homogeneous, taxoid C-ring C-10 electrophiles **5**, **6** and C-10 nucleophiles **18**, **19** that offer distinct linking possibilities with various A-ring precursors is described. The key feature of the synthetic scheme reported in this paper is the lead tetraacetate mediated consecutive domino reaction that enables the transformation of fused bicyclic diol **1** into bridged bicyclic

aldol **7** in a single synthetic operation. Our retrosynthetic analysis calls for an introduction of the missing C-2 carbon in a post-coupling operation where an alkoxymethylenation would provide the electrophilic partner in the C-1–C-2 bonding.

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Introduction

Three out of six total syntheses of taxol^[1] used a convergent A+C approach; in such an approach, two six-membered ring fragments have to be constructed, coupled and elaborated to form the taxoid diterpene skeleton. Nicolaou, [1a] in his total synthesis of taxol, by proceeding in the A+C direction, constructed the C-ring precursor starting from mucic acid, whereas Danishefsky synthesized a fully functional C-ring starting from (S)-(+)-Wieland-Miescher ketone.^[1c] Finally, Kuawajima^[1f] started the synthesis of his C-ring precursor from 2-bromocyclohexenone, which was converted in eight steps into a cyclohexadiene derivative and used in the segment coupling. Recent papers from these laboratories have described a route to taxoid C-ring systems of type 2 that involved a Pb(OAc)₄ mediated domino^[2] reaction (oxidative cleavage/cycloaddition/oxymetallation/ring expansion) of (S)-(+)-Hajos-Parrish ketone derived unsaturated diols of type 1 (Scheme 1). Our previous work, a convergent A+C approach to the taxoid ABC framework with a Still α -alkoxylithium methodology^[3] (top side linking) and an intramolecular aldol reaction (bottom side linking) as key bond constructions, had demonstrated the suitability of the domino chemistry to access ABC-taxoid diterpene skeleton 3 (longest linear sequence: eight steps).^[4] Although this method was reliable, we investigated an alternative route, which would produce the desired products more briefly without recourse to post-coupling operations at the C-ring level.



Scheme 1.

Even though a retrosynthetic analysis to construct a pentafunctionalized six-membered ring containing two quaternary centers may seem an easy task, severe length and selectivity problems could be encountered towards its realization. Our desire to improve the overall efficiency of this process and expand its applicability to related ring systems prompted us to examine modifications to this sequence. Despite the anticipated difficulty, on the basis of the proposed mechanism^[5] of carrying out a domino reaction with a hindered unsaturated bicyclic diol such as 1b we first investigated the latter as a model system containing the C-5 αhydroxy group that is necessary for esterification. [6] To secure the formation of domino product 2b, diol 1b was treated with lead tetraacetate in a preheated oil bath at 100 °C (instead of the usual 25 °C) and provided the desired domino product in 21% isolated yield; far too inefficient to be synthetically useful because of the low mass recovery and low purity of the crude reaction product. Notwithstanding the high degree of oxygenation of 2b thus obtained, its synthetic utility was compromised and so we had

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to seek an alternative strategy. The synthetic route chosen for the construction of the taxoid C-ring is based on the recently developed building block approach to bicyclo[2.2.2]aldol 7a (Scheme 2) which utilizes a more benign version of the Pb(OAc)₄ induced heterodomino transformations with the use of only 1 equiv. of lead tetraacetate.[7] This three-reagent consecutive domino reaction initiated by PhI(OAc)₂ (oxidative/pericyclic), continued by Pb(OAc)₄ (ring expansion), and completed by a mild base consisting of solid K₂CO₃ (ring system interchange) all in one-pot allowed for a large scale preparation of key intermediate 7a. This bicyclic aldol framework offered chemoselectivity because of the threefold different oxygenation pattern, and stereoselectivity, given that chemical modifications of bridged polycyclic compounds are highly stereoselective. Thus, with the aim of minimizing post-coupling elaboration, we focused on 11a as an exact embodiment of a taxoid right-half moiety (C-ring), which contains all of the stereogenic centers in the desired absolute configuration and functionality at C-3/C-10 for linking with the A-ring counterpart. Our new goal, therefore, became the generation of a more functionalized C-ring candidate of type 5 or 6, prior to C-10-C-11 linkage (taxoid numbering used throughout the text). Described below is an efficient route starting from bicyclic diol 1a, which was converted to heavily substituted cyclohexane derivatives of type 5, 6, 18, or 19 through a consecutive domino reaction followed by oxidation ring opening maneuvers (Schemes 3, 4, 5 and 6).

Results and Discussion

The taxoid C-ring has to contain a functional moiety that could ultimately be used to introduce the C-5 α-OH and the C-20 carbon that would offer further functionality; this would therefore ensure access to oxirane-, oxetane-, or olefin-containing taxoid families. The synthesis started with the experimentally simple reactions employed in effecting the one-pot transformation of fused bicyclic diol 1a to bridged bicyclic aldol 7a (Scheme 2) and hence to its TBSprotected derivative 7b in large scale by using the protocol published in ref.^[7] Two straightforward steps were then required for the incorporation of the oxygen functionalities at C-3 and the C-20 hydroxymethyl substituent. The best conversion of 7b into requisite acyloin 8 was achieved by the anomalous ozonization of the silvl enol ether derivative of **7b** (MeOH, –78 °C) as described in the preceding article. This resulted in a 77.4% combined yield of a 21.7:1 mixture of endo-8 and its corresponding exo-8 adduct, separated by chromatography. On the other hand, direct acetoxylation of 7b by treatment with Pb(OAc)₄ in benzene (90 °C, 2 d) afforded a modest 35% isolated yield of required α-acetoxyacyloin 8 along with unreacted starting material (40%). Subsequent addition of the α -alkoxyorganolithium derived from Bu₃SnCH₂OMOM^[8] by transmetalation with nBuLi (nBuLi, Bu₃SnCH₂OMOM, in dry THF, -78 °C, 30 min) gave a mixture of two products: 9 and its deacetylated counterpart in a 75% combined yield and 1:2.7 ratio, together with recovered starting material (10%). All were separated by SiO₂ flash chromatography and isolated pure; the deacetylated product was quantitatively recycled by reacetylation (Ac₂O, pyridine, DMAP). Compound 9 was desilylated with tetrabutylammonium fluoride (TBAF, THF, 0 °C, 91%) to afford the corresponding alcohol which was further oxidized with the Dess-Martin protocol to required ketone 4 in nearly quantitative yield (Dess-Martin periodinane, CH₂Cl₂, pyridine, 25 °C, 96%).

Scheme 2. a) 1.2 equiv. PhI(OAc)₂, 24 h, then 1.2 equiv. Pb(OAc)₄, PhMe, 15 h; then K₂CO₃-MeOH, H₂O, 25 °C; b) TBSOTf, collidine, CH₂Cl₂, 0 °C, 30 min; c) Pb(OAc)₄, PhH, 90 °C, 48 h; d) Bu₃SnCH₂OMOM, nBuLi, -78 °C; e) TBAF, THF, 0 °C; f) DMP, pyridine, CH₂Cl₂, 25 °C, 1 h; g) m-CPBA, NaHCO₃, CH₂Cl₂, 25 °C.

It was expected that Baeyer-Villiger oxidations of bridged bicyclic aldol 4 could afford lactones with reasonable control of bridgehead migration over methylene migration. In light of the intended application of this approach to the construction of taxoid C-ring precursors 5 and 6, this concern had to be addressed. It thus turned out to be our initial objective to demonstrate that bicyclo[2.2.2] aldol 4 could be transformed into corresponding lactone 11a; reductive or hydrolytic opening of the latter would in turn allow access to the desired C-ring candidates. To this end, 4 was oxidized with excess m-chloroperbenzoic acid in sodium hydrogencarbonate buffered dichloromethane, at room temperature to lactones 10 and 11a, which result from methylene and bridgehead migration, respectively, in 60% isolated yield and a 10/11a ratio of 1:4, along with recovered starting material (35%). The isomers were separated by chromatography and the major product was identified as lactone 11a. The course of the Baeyer-Villiger oxidation^[9] of **4** was readily determined, as the regiochemistry of the oxygen insertion was unambiguously confirmed by long-range heteronuclear couplings observed in the HMBC spectra. Ratios of the bridgehead to methylene migrated lactones were determined by SiO₂ separation and by comparison of the integrated areas for the methylene protons that are adjacent to the carbonyl group in bridgehead-migrated lactone 11a (at δ = 2.24 and 2.52 ppm) to the oxygen in methylene-migrated lactone 10 ($\delta = 3.74$ and 4.58 ppm) and the bridgehead proton at $\delta = 3.18$ ppm in 10 which is absent in 11a. Interestingly, this oxidation, unlike bicyclo[2.2.2]octanone counterpart 7a, [6] gave a significant

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amount of isomeric lactone 10. At this stage it was decided to investigate procedures to improve both the yields and the 11a/10 ratio. On the basis of precedent, it seemed likely that the nature of the peroxy acid used may influence the results obtained. Among the various oxidants tried, including tBuO₂H, NaOH, THF,^[10] NaBO₃, AcOH,^[11] MMPP, DMF,[12] H₂O₂, AcONa, AcOH,[13] PhSeO₂H, H₂O₂, CH₂Cl₂, and H₂O,^[14] none led to either one of the lactones; the only new compound was the C-3 deacetylated starting material when basic conditions were used (tBuO₂H-NaOH). The Sc(OTf)₃ or Yb(OTf)₃ catalyzed^[15] Baeyer– Villiger oxidation with m-CPBA in CH₂Cl₂ was also tried but lead only to some tBu-deprotection and poor mass balance. The reactions were run under variable conditions. The temperature, number of equivalents of the oxidizing reagent, reaction times, and the nature of the oxidant used were all varied. Microbial Baeyer-Villiger oxidation^[16] was also tried with the use of the fungus Cunninghamella echinulata[17] but failed to produce any lactone in a detectable amount. To summarize, while all attempts to increase the yields of lactones 10/11a were unsuccessful, we were happy with an isolated 60% yield because the unreacted starting material (35%) was easily recovered by chromatography because of its large $R_{\rm f}$ difference and could therefore be recycled. Also, despite efforts to minimize the formation of undesired lactone 10, the product distribution remained in the range of 4:1. One of the main points about the bridge migration during Baeyer-Villiger oxidation of bicyclic aldol 4 was that lactone i (oxabicyclo[3.2.2]nonan), initially formed as an electronically controlled transition state, is an intermediate in the formation of lactone 11a (oxabicyclo[3.3.1]nonan) towards which it evolves by spontaneous translactonization (Scheme 3).

The illustrated stereochemistries at the newly installed stereogenic centers, as well as bond connectivities, within these products follow from extensive NMR spectroscopic

Scheme 3.

studies and were further corroborated by an X-ray crystallographic study on 10 and 11a (Figure 1).

With lactone 11a available, its conversion to 14 was explored (Scheme 4). TBS protection^[18] of the free hydroxy group at C-5 (TBSOTf, collidine, CH₂Cl₂, 0 °C, 30 min) led to 11b (Scheme 2), together with orthoester 11c (95% combined yield, 4.3:1 ratio), which upon fluoride deprotection (TBAF, THF, 0 °C), was converted to 11a and recycled. Reductive opening of the lactone was first accomplished by treatment with excess LiAlH₄, in THF at 0 °C. These conditions caused partial cleavage of the TBS ether to afford desired triol 12 (52%) accompanied by tetraol 13 (30%) and fused bicyclic lactone 5 (11%), which results from partial saponification at C-3 and subsequent intramolecular translactonization. Triol 12 was then converted into target Cring C-10 electrophile 6 in two straightforward steps. Selective acetonide formation (acetone, p-TsOH, 25 °C, 3 h) furnished desired isopropylidene alcohol 14 (84% isolated yield), which in turn was oxidized with the use of the Dess-Martin protocol (DMP, pyridine, CH₂Cl₂, 25 °C, 30 min) to afford 6 (86%). Tetraol 13 could also be obtained by direct reduction of C-5-unprotected lactone 11a under the same conditions. Subsequent selective acetonide formation (acetone, p-TsOH, 0 °C, 2.5 h) furnished 15 in 80% isolated yield. The latter can be used in the synthesis due to easy differentiation of the C-5/C-10 positions. The synthesis of a

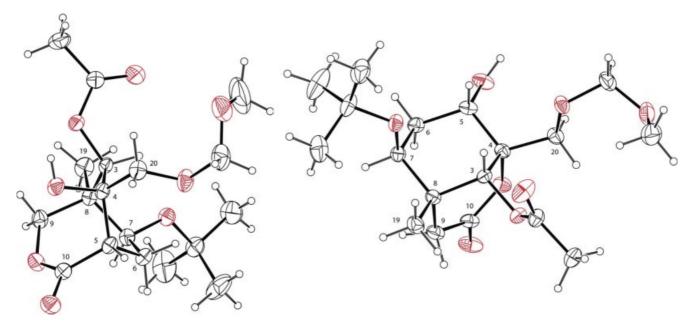


Figure 1. X-ray structures of 10 and 11a.

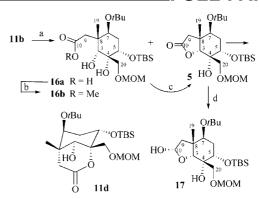
fully functional taxoid C-ring precursor, C-10 electrophile **6**, which offers linking possibilities, is portrayed in Scheme 4.

Scheme 4. a) LiAlH₄, THF, 0 °C, 2 h; b) acetone, *p*-TsOH, 25 °C, 2.5 h; c) Dess–Martin periodinane, CH₂Cl₂, pyridine, 25 °C, 0.5 h.

As mentioned above, reductive lactone opening was somehow complicated by loss of the tert-butyldimethylsilyl group which could reach levels as high as 30% depending upon reaction time and work up conditions. Other reducing agents were screened for transformation of 11b into target molecule 6, though without any significant improvement. To obviate these complications, we turned our attention to a hydrolytic lactone opening with K₂CO₃/MeOH-H₂O. Subjecting 11b to these conditions for 20 h at room temperature gave a 90% combined yield of 16a and 5 in a 4:1 ratio, respectively. Compound 16a, which shows a marked tendency to lactonize on standing, was esterified (TMSCHN₂, Et₂O, MeOH, 25 °C, 1 h) to afford a 90% yield of 16b, which again showed the same tendency towards formation of lactone 5. Nearly quantitative formation of lactone 5 was secured while attempting to form the corresponding isopropylidene derivative (p-TsOH, acetone, 25 °C, 12 h).

As portrayed in Scheme 5, during the conversion of bridged bicyclic lactone 11b to desired cyclohexane 16 upon hydrolytic lactone opening, 11d is formed initially but rapidly converts to fused bicyclic lactone 5 by an intramolecular transesterification to give a less sterically congested product. This was proven experimentally by isolating and identifying 11d during conversion. DIBAL reduction of 5 (DIBAL, PhMe, -78° to -40 °C, 2 h) gave a 93% isolated yield of lactol 17 along with trace amounts of its epimer (not isolated pure). Single-crystal X-ray diffraction analysis of crystalline lactol 17 (Figure 2), enabled unequivocal stereochemical assignment and further confirmed the correctness of previous assignments by NMR techniques throughout the synthetic scheme.

Finally, a two-step umpolung sequence applied to taxoid C-ring C-10 electrophile 6 then afforded α -alkoxyorganostannanes 18 and 19 to be used as the C-10 nucleophile precursors in our A+C approach (Scheme 6). Treatment of 6 with lithium tributylstannylate (in dry THF, -78 °C, 20 min) followed by protection of the resulting alcohol with chloromethyl methyl ether in the presence of Hünig's base (iPr_2NEt) gave a mixture of two products, 18 and 19 (75%



Scheme 5. a) $K_2CO_3/MeOH/H_2O$, 25 °C; b) TMSCHN₂, Et₂O, MeOH, 25 °C, 1 h; c) *p*-TsOH, acetone, 25 °C, 12 h; d) DIBAL, PhMe, -78° to -40 °C, 2 h.

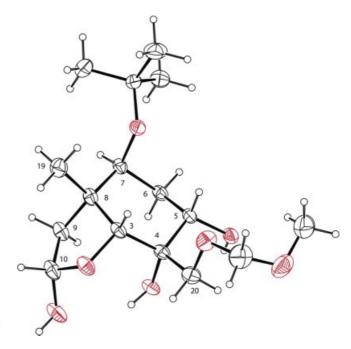


Figure 2. X-ray structure of 17.

in 1:1 ratio), which were separated by chromatography and isolated pure. The two α -alkoxyorganostannanes thus obtained will herein be arbitrarily referred to as higher- R_f 18 and lower- R_f 19.

Scheme 6. a) Bu₃SnH, LDA, THF, -78 °C; then MOMCl, iPr_2Net , CH₂Cl₂, 25 °C.

This completes the synthesis of second generation taxoid C-ring precursors. This process is considerably improved over the previously published routes. The observation that Still coupling is a useful reaction for the union of subunits **20** and **21** (first generation taxoid A- and C-rings, respectively) justifies the use of this reaction sequence in convergent strategies. By preparing compounds **5**, **6**, **18**, and **19** an additional functionalization step (the C-5 α -hydroxy function) was inserted prior to the beginning of the linear synthesis, while the C-20 hydroxymethyl group at the quaternary C-4 center was already installed.

Conclusions

From a practical standpoint, unraveling the bicyclo-[2.2.2]octane moiety in a single step from bicyclic diol 1a, rapidly introducing the C-3 then C-20 substituents and subjecting it to Baeyer–Villiger conditions offered the opportunity to prepare the second generation C-ring precursors. The most critical issues of our synthesis were the *m*-CPBA mediated Baeyer–Villiger oxidation and subsequent reductive lactone ring opening. Although the yields of these conversions were far from excellent, practical access to the required substrates became available. Overall, the approach represents an interesting application of LTA-mediated domino methodology, which provides fully functional, optically pure C-ring precursors minimizing post-coupling elaboration and nicely complements the C-ring syntheses published to date.

Experimental Section

General experimental details were previously described. [7] The term "usual work up" refers to washing the organic layer with brine, drying with anhydrous MgSO₄, and evaporating the solvent in vacuo with a rotary evaporator at aspirator pressure. Experimental evidence favoring the structures came from a comprehensive range of ¹H- and ¹³C NMR spectroscopic data (1D and 2D experiments) and corroborated by spatial proximity (n.O.e) studies. X-ray analysis of 10, 11a, and 17 supports the suggested structures. CCDC-618901 (10), -618902 (11a), and -618903 (17) contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

Elaboration of the C-4 Quaternary Center – Preparation of 9: To a magnetically stirred solution of Bu₃SnCH₂OMOM (6.46 g, 17.6 mmol) in anhydrous THF (15 mL) cooled to –78 °C under an argon atmosphere, *n*BuLi (1.6 m in hexane, 10.8 mL, 17.2 mmol) was added, and the mixture was stirred at this temperature for 10 min before **8** (2.93 g, 7.37 mmol) was added. After stirring 30 min at –78 °C, the reaction mixture was diluted with ethyl acetate and quenched with a saturated solution of NH₄Cl. Following usual work up, SiO₂ column chromatography (heptane/EtOAc, 15:1 to EtOAc) afforded 699 mg (20%) of **9**, 1.75 g (55%) of C-3-saponified diol **9**, along with recovered starting material **8** (295 mg, 10%).

7-*tert*-Butoxy-5-(*tert*-butyldimethylsilanyloxy)-3-hydroxy-3-methoxymethoxymethyl-1-methylbicyclo[2.2.2]oct-2-yla Acetate (9): $[a]_D^{20}$ = +47 (c 1.0, CHCl₃). M.p. 137–139 °C (CH₂Cl₂). IR (film): \tilde{v} = 3451, 2973, 2953, 2934, 2858, 1731, 1247, 1046 cm⁻¹. ¹H NMR

(300 MHz, CDCl₃): δ = 0.08 (s, 3 H), 0.10 (s, 3 H), 0.87 (s, 3 H), 0.90 (s, 9 H), 1.13 (s, 9 H), 1.53 (ddd, J = 1.7, 9.5, 14.7 Hz, 1 H), 1.62 (ddd, J = 2.5, 8.8, 15.0 Hz, 1 H), 1.74 (ddd, 1 H, J = 2.3, 3.9, 15.0 Hz, 1 H), 1.82 (dd, J = 2.3, 14.7 Hz, 1 H), 2.07 (s, 3 H), 2.15 (q, J = 3.5 Hz, 1 H), 3.22 (dd, J = 2.2, 8.8 Hz, 1 H), 3.39 (s, 3 H), 3.60 (d, J = 10.9 Hz, 1 H), 4.00 (ddd, J = 2.3, 3.7, 9.5 Hz, 1 H), 4.09 (dd, J = 2.7, 10.9 Hz, 1 H), 4.54 (d, J = 1.5 Hz, 1 H), 4.69 (s, 2 H), 4.95 (d, J = 2.5 Hz, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = -5.1, -4.9, 17.7, 20.9, 21.3, 25.7, 28.6, 33.0, 36.5, 37.0, 37.3, 55.4, 71.0, 71.1, 73.2, 73.3, 73.5, 97.2, 170.9 ppm. ESIMS (MeOH): m/z (%) = 497.2 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for $C_{24}H_{46}O_7NaSi$ 497.2911, found 497.2916. $C_{24}H_{46}O_7Si$ (474.30): calcd. C 60.72, H 9.77; found C 60.89, H 10.56.

7-tert-Butoxy-5-(tert-butyldimethylsilanyloxy)-3-methoxymethoxymethyl-1-methylbicyclo[2.2.2]octane-2,3-diol: $[a]_D^{20} = +52$ (c 1.6, CHCl₃). M.p. 42–43 °C (CH₂Cl₂). IR (film): $\tilde{v} = 3394$, 2956, 2930, 2860, 1043, 839 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.09$ (2s, 6 H), 0.90 (s, 9 H), 0.93 (s, 3 H), 1.11 (s, 9 H), 1.48 (ddd, J = 1.7, 9.7, 14.5 Hz, 1 H), 1.56 (dt, J = 3.3, 14.9 Hz, 1 H), 1.74 (ddd, J =3.2, 9.3, 14.9 Hz, 1 H), 1.79 (dd, J = 4.4, 14.5 Hz, 1 H), 2.10 (q, J= 3.2 Hz, 1 H), 3.23 (dd, J = 3.4, 9.3 Hz, 1 H), 3.39 (s, 3 H), 3.41 Hz(d, J = 1.7 Hz, 1 H), 3.46 (s, 1 H), 3.50 (d, J = 10.8 Hz, 1 H), 3.70(dd, J = 1.7, 10.8 Hz, 1 H), 3.95 (ddd, J = 3.0, 4.4, 9.7 Hz, 1 H),4.67 (d, J = 6.5 Hz, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta =$ -5.2, -4.8, 17.8, 20.4, 25.7, 28.6, 33.6, 36.0, 37.2, 38.4, 55.3, 68.4, 70.6, 71.7, 72.4, 73.1, 74.2, 96.9 ppm. ESIMS (MeOH): m/z (%) = 455.2 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for C₂₂H₄₄O₆SiNa 455.2805; found 455.2812. C₂₂H₄₄O₆Si (432.29): calcd. C 61.07, H 10.25; found C 61.08, H 10.24.

Preparation of the Baeyer–Villiger Precursor – Bicyclic Ketone 4: To a magnetically stirred solution of 9 (1.64 g, 3.46 mmol) in dry THF (35 mL) at –75 °C was added tetrabutylammonium fluoride (1 m in tetrahydrofuran, 10.4 mmol). The reaction mixture was stirred and warmed from –75 °C to –45 °C for 1 h 30 min (TLC monitoring). Ethyl acetate was then added, and following extraction the crude mixture was worked up as usual and purified by silica gel column chromatography (heptane/EtOAc, 3:1 to EtOAc) to give the diolacetate (1.14 g, 91%).

7-tert-Butoxy-3,5-dihydroxy-3-methoxymethoxymethyl-1-methylbi**cyclo[2.2.2]oct-2-yl** Acetate: $[a]_D^{20} = +75$ (c 1.1, CHCl₃). M.p. 99– 100 °C (CH₂Cl₂). IR (film): $\tilde{v} = 3396, 2973, 2934, 1732, 1455, 1368,$ 1247, 1046 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.84$ (s, 3 H), 1.13 (s, 9 H), 1.57 (td, J = 3.3, 14.9 Hz, 1 H), 1.65 (ddd, J = 1.8, 9.8, 14.7 Hz, 1 H), 1.74 (ddd, J = 3.0, 9.2, 14.9 Hz, 1 H), 1.84 (dd, J = 3.9, 14.7 Hz, 1 H), 2.11 (s, 4 H), 2.13 (m, 1 H), 3.26 (dd, J =3.0, 9.2 Hz, 1 H), 3.39 (s, 3 H), 3.66 (d, J = 10.9 Hz, 1 H), 3.87 (td, J = 10.9 Hz) J = 3.6, 9.8 Hz, 1 H), 3.94 (br. s, 1 H), 3.96 (d, J = 10.9 Hz, 1 H), 4.63 (d, J = 6.5 Hz, 1 H), 4.67 (d, J = 6.5 Hz, 1 H), 4.74 (d, J =1.8 Hz, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 20.6, 20.9, 28.7 (3 C), 33.8, 37.3, 37.5, 38.1, 55.6, 70.0, 70.8, 72.2, 73.5, 74.3, 74.5, 97.4, 170.9 ppm. ESIMS (MeOH): *m/z* (%) = 383.2 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for C₁₈H₃₂O₇Na 383.2046; found 383.2061. C₁₈H₃₂O₇ (360,21): calcd. C 59.98, H 8.95; found C 60.07, H 8.95.

7-tert-Butoxy-3-hydroxy-3-methoxymethoxymethyl-1-methyl-5-oxobicyclo[2.2.2]oct-2-yl Acetate (4): A dry flask was charged with starting diols (1.13 g, 3.14 mmol) in dichloromethane (28 mL), pyridine (2.5 mL, 28.3 mmol), and Dess-Martin periodinane (3.62 g, 8.54 mmol). The mixture was stirred at room temperature for 1 h, (TLC monitoring). The reaction was then diluted with dichloromethane, quenched with a saturated aqueous solution of sodium hydrogencarbonate, and worked up as usual to afford, after purifi-

cation on silica gel column chromatography (heptane/EtOAc 2:1 to EtOAc), 1.08 g (96%) of 4.[a] $_{0}^{20}$ = +71 (c 1.2, MeOH). M.p. 92–93 °C (CH $_{2}$ Cl $_{2}$). IR (film): \bar{v} = 3446, 2972, 1735, 1373, 1232, 1055 cm $^{-1}$. 1 H NMR (300 MHz, CDCl $_{3}$): δ = 0.92 (s, 3 H), 1.12 (s, 9 H), 1.73 (td, J = 3.0, 15.2 Hz, 1 H), 1.81 (dd, J = 1.7, 19.2 Hz, 1 H), 2.02 (ddd, J = 3.0, 9.4, 15.2 Hz, 1 H), 2.06 (s, 3 H), 2.54 (d, J = 19.1 Hz, 1 H), 2.54 (t, J = 3.1 Hz, 1 H), 3.04 (s, 1 H), 3.33 (s, 3 H), 3.45 (dd, J = 3.0, 9.3 Hz, 1 H), 3.74 (d, J = 11.0 Hz, 1 H), 3.91 (d, J = 11.0 Hz, 1 H), 4.57 (d, J = 6.5 Hz, 1 H), 4.61 (d, J = 6.5 Hz, 1 H), 4.93 (d, J = 1.9 Hz, 1 H) ppm. 13 C NMR (75 MHz, CDCl $_{3}$): δ = 19.7, 20.7, 28.5 (3 C), 31.6, 41.1, 42.4, 51.4, 55.5, 70.4, 71.0, 72.4, 73.4, 73.8, 97.2, 170.7, 211.5 ppm. ESIMS (MeOH): m/z (%) = 381.1 (100) [M + Na] $^+$. HR ESIMS (MeOH): calcd. for C $_{18}$ H $_{30}$ O $_{7}$ Na 381.1889; found 381.1854. C $_{18}$ H $_{30}$ O $_{7}$ (358.20): calcd. C 60.32, H 8.44; found C 60.27, H 8.52.

Baeyer–Villiger Oxidation of 4: Sodium hydrogenearbonate (642 mg, 7.64 mmol) and *m*-chloroperbenzoic acid (3.30 g, 19.10 mmol) were added to **4** (1.14 g, 3.18 mmol) in dichloromethane (20 mL) at 0 °C. The mixture was stirred at room temperature for approximately 17 h (TLC monitoring). The crude reaction mixture was diluted with dichloromethane and filtered through a plug of Celite. The excess peracid was decomposed by washing with aqueous 5% sodium sulfite. Finally, the organic phase was washed with saturated NaHCO₃ and worked up as usual to afford, after purification by flash chromatography (CH₂Cl₂/Me₂C=O, 95:5), 398 mg (35%) of starting material, 142 mg (12%) of **10**, and 571 mg (48%) of **11a**. For large scale experiments the lactone separation is better carried out after the TBS-protection step.

9-tert-Butoxy-7-hydroxy-7-methoxymethyl-5-methyl-2-oxo-**3-oxabicyclo**[3.2.2]non-6-yl Acetate (10): $[a]_D^{20} = +109$ (c 0.3, CHCl₃). M.p. 113–114 °C (CH₂Cl₂). IR (film): $\tilde{v} = 3446$, 2972, 1735, 1373, 1232, 1055 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ = 0.83 (s, 3 H), 1.18 (s, 9 H), 1.72 (ddd, J = 4.4, 6.0, 15.3 Hz, 1 H), 2.15 (s, 3 H), 2.42 (ddd, J = 3.0, 8.8, 15.3 Hz, 1 H), 3.18 (dd, J =3.1, 4.3 Hz, 1 H), 3.34 (s, 1 H), 3.39 (s, 3 H), 3.57 (d, J = 10.5 Hz, 1 H), 3.62 (d, J = 10.5 Hz, 1 H), 3.74 (dd, J = 1.5, 12.2 Hz, 1 H), 3.84 (dd, J = 6.0, 8.8 Hz, 1 H), 4.58 (d, J = 12.2 Hz, 1 H), 4.61 (d, J = 12.2 Hz, 1 H)J = 6.5 Hz, 1 H), 4.64 (d, J = 6.5 Hz, 1 H), 5.11 (d, J = 1.5 Hz, 1 Hz) H) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 18.8, 20.9, 28.7 (3 C), $32.3,\,42.5,\,48.8,\,55.8,\,68.6,\,71.1,\,71.9,\,72.0,\,73.7,\,74.1,\,97.4,\,170.2,$ 172.9 ppm. ESIMS (MeOH): m/z (%) = 397.1 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for C₁₈H₃₀O₈Na 397.1838; found 397.1830. C₁₈H₃₀O₈ (374.19): calcd. C 57.74, H 8.08; found C 57.77, H 8.07.

6-tert-Butoxy-8-hydroxy-1-methoxymethoxymethyl-5-methyl-3-oxo-2-oxabicyclo]3.3.1]non-9-yl Acetate 11a: $[a]_D^{20} = +44$ (c 1.6, CHCl₃). M.p. 143–145 °C (CH₂Cl₂). IR (film): $\tilde{v} = 3454$, 2973, 1737, 1372, 1228, 1036 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.95$ (s, 3 H), 1.20 (s, 9 H), 1.46 (ddd, J = 2.6, 11.9, 14.5 Hz, 1 H), 2.12 (s, 3 H), 2.16 (ddd, J = 2.8, 5.2, 14.5 Hz, 1 H), 2.28 (d, J = 19.0 Hz, 1 H), 2.54 (d, J = 19.0 Hz, 1 H), 3.35 (s, 3 H), 3.36 (d, J = 8.6 Hz, 1 H), 3.44 (t, J = 2.7 Hz, 1 H), 3.60 (d, J = 9.8 Hz, 1 H), 3.82 (d, J = 9.8 Hz, 1 H), 4.23 (ddd, J = 5.2, 8.6, 11.5 Hz, 1 H), 4.58 (d, J = 6.5 Hz, 1 H), 4.64 (d, J = 6.5 Hz, 1 H), 5.38 (s, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta = 20.6$, 22.5, 28.7 (3 C), 34.0, 38.2, 39.8, 55.6, 65.4, 67.0, 67.6, 72.9, 74.7, 85.3, 97.1, 169.4, 169.7 ppm. ESIMS (MeOH): m/z (%) = 397.1 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for C₁₈H₃₀O₈Na 397.1838; found 397.1804. C₁₈H₃₀O₈ (374.19): calcd. C 57.74, H 8.08; found C 57.59, H, 8.01.

Baeyer–Villiger Oxidation of **4** with *t*BuO₂H as the oxidant yielded only deacetylated **4**. A mixture of aldol **4** (51.3 mg, 0.14 mmol), *tert*-butyl hydroperoxide (70% aq., 55 μL, 0.43 mmol), and NaOH

(10% aq., 68 μL, 0.17 mmol) in THF (1.4 mL) cooled to 0 °C was stirred for 30 min, and the mixture was then warmed to room temperature and stirred for ca. 6 h (TLC monitoring). The crude reaction mixture was taken up in ether (20 mL), the excess peracid was decomposed by washing with aqueous 5% sodium sulfite, and the organic phase was worked up as usual to give, after purification by silica gel column chromatography (heptane/AcOEt, 2:1 to AcOEt), only deacetylated starting material (C-3-OH)-4 [41.6 mg (81%)].

8-tert-Butoxy-5,6-dihydroxy-6-methoxymethoxymethyl-4-methylbicyclo[2.2.2]octan-2-one: $[a]_{20}^{20} = +72$ (c 0.9, CHCl₃). IR (film): $\tilde{v} = 3442$, 2973, 2933, 1726, 1366, 1075, 1038 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 1.04$ (s, 3 H), 1.15 (s, 9 H), 1.65 (ddd, J = 2.7, 3.8, 15.1 Hz, 1 H), 1.75 (dd, J = 2.1, 19.1 Hz, 1 H), 2.07 (ddd, J = 3.6, 9.4, 15.1 Hz, 1 H), 2.52 (d, J = 19.1 Hz, 1 H), 2.53 (dd, J = 2.7, 3.4 Hz, 1 H), 3.1 (d, J = 6.4 Hz, 1 H), 3.41 (s, 3 H), 3.45 (dd, J = 3.8, 9.4 Hz, 1 H), 3.63 (d, J = 11.0 Hz, 1 H), 3.77 (dd, J = 1.9, 6.0 Hz, 1 H), 3.88 (d, J = 11.0 Hz, 1 H), 3.99 (s, 1 H), 4.66 (d, J = 6.6 Hz, 1 H), 4.69 (d, J = 6.6 Hz, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta = 19.5$, 28.6 (3 C), 32.0, 42.2 (2 C), 51.4, 55.8, 68.2, 70.7, 72.0, 73.0, 73.7, 97.5, 212.1 ppm. ESIMS (MeOH): m/z (%) = 339.1 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for C₁₆H₂₈O₆Na 339.1784; found 339.1788. C₁₆H₂₈O₆ (319.19) ·0.7H₂O: calcd. C 58.41, H 9.01; found C 58.43, H 8.75.

TBS-protection of C-5-OH: To a stirred solution of 11a (163 mg, 0.44 mmol) and collidine (0.17 mL, 1.31 mmol) in dry dichloromethane (4.0 mL) at 0 °C under an argon atmosphere, was added TBSOTf (0.20 mL, 0.87 mmol). The mixture was stirred at room temperature for 30 min (TLC monitoring), diluted with heptane, washed with diluted NaHCO₃ and worked up as usual. After purification by silica gel column chromatography (heptane/AcOEt, 5:1 to AcOEt), 163 mg (77%) of 11b, and 39 mg (18%) of secondary product 11c were obtained.

6-tert-Butoxy-8-(tert-butyldimethylsilanyloxy)-1-methoxymethoxymethyl-5-methyl-3-oxo-2-oxabicyclo[3.3.1]non-9-yl Acetate (11b): $[a]_{D}^{20} = +6$ (c 0.8, CHCl₃). M.p. 89–90 °C (CH₂Cl₂). IR (film): $\tilde{v} =$ 2930, 2857, 1741, 1228, 1116, 1038, 837 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.06$ (s, 3 H), 0.07 (s, 3 H), 0.88 (s, 9 H), 0.94 (s, 3 H), 1.21 (s, 9 H), 1.55 (ddd, J = 2.6, 11.7, 14.3 Hz, 1 H), 1.99 (ddd, J= 3.0, 5.0, 14.3 Hz, 1 H), 2.12 (s, 3 H), 2.24 (d, J = 19.0 Hz, 1 H),2.52 (d, J = 19.0 Hz, 1 H), 3.33 (s, 3 H), 3.40 (t, J = 2.7 Hz, 1 H), $3.59 \text{ (d, } J = 9.1 \text{ Hz, } 1 \text{ H), } 3.72 \text{ (d, } J = 9.1 \text{ Hz, } 1 \text{ H), } 4.26 \text{ (dd, } J = 9.1 \text{$ 5.0, 11.7 Hz, 1 H), 4.54 (d, J = 6.6 Hz, 1 H), 4.59 (d, J = 6.6 Hz, 1 H), 5.39 (s, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta = -4.8$, -4.1, 18.0, 20.7, 22.7, 25.8 (3 C), 28.7 (3 C), 34.6, 38.4, 39.8, 55.9, 64.6, 66.9, 67.7, 73.2, 74.5, 85.6, 97.3, 169.5, 169.8 ppm. ESIMS (MeOH): m/z (%) = 511.2 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for $C_{24}H_{44}O_8NaSi\ 511.2703;$ found 511.2669. $C_{24}H_{44}O_8Si$ (488.28): calcd. C 58.99, H 9.08; found C 59.16, H, 9.03.

5-*tert*-**Butoxy-1-**(*tert*-**butyldimethylsilanyloxy**)-**8-**methoxymethoxymethyl-**6-**methyl-**2,9-**dioxatricyclo[**4.1.1.1**^{1.6}]dec-7-yl Acetate (11c): $[a]_{20}^{20} = +61$ (c 0.5, CHCl₃). IR (film): $\tilde{v} = 2929$, 2857, 1746, 1367, 1232, 1039, 840 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.18$ (s, 6 H), 0.84 (s, 3 H), 0.91 (s, 9 H), 1.15 (s, 9 H), 1.54 (ddd, J = 0.9, 7.5, 14.7 Hz, 1 H), 1.69 (d, J = 13.6 Hz, 1 H), 2.00 (d, J = 13.6 Hz, 1 H), 2.14 (s, 3 H), 2.51 (ddd, J = 4.2, 8.1, 14.7 Hz, 1 H), 3.35 (s, 3 H), 3.47 (d, J = 10.0 Hz, 1 H), 3.57 (d, J = 10.0 Hz, 1 H), 3.71 (t, J = 7.8 Hz, 1 H), 4.19 (dd, J = 0.9, 4.2 Hz, 1 H), 4.58 (d, J = 6.5 Hz, 1 H), 4.63 (d, J = 6.5 Hz, 1 H), 5.15 (s, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta = -3.4$, -3.3, 17.8, 20.7, 20.9, 25.8 (3 C), 28.7 (3 C), 35.5, 39.6, 46.8, 55.4, 67.6, 68.3, 73.3, 73.6, 74.6, 80.6, 97.0, 117.1, 170.7 ppm. ESIMS (MeOH): m/z (%) = 511.2 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for C₂₄H₄₄O₈NaSi

511.2703; found 511.2697. $C_{24}H_{44}O_8Si$ (488.28): calcd. C 58.99, H 9.08; found C 58.82, H 9.03.

Reductive Opening of Lactone 11b with Lithium Aluminum Hydride and Selective Acetonide Formation: To a magnetically stirred suspension of LiAlH₄ (24 mg, 0.64 mmol) in THF (2 mL) at 0 °C was added a solution of 11b (156 mg, 0.32 mmol) in THF (2.5 mL). The mixture was stirred at this temperature for 1 h (TLC monitoring), diluted with diethyl ether, worked up as usual to give, after purification on silica gel column chromatography (heptane/AcOEt, 5:1 to AcOEt) 15.7 mg (11%) of 5, 75 mg (52%) of 12, along with its TBS-deprotected counterpart 13 (32 mg, 30%).

4-tert-Butoxy-6-(tert-butyldimethylsilanyloxy)-3-(2-hydroxyethyl)-1methoxymethyl-3-methyl-cyclohexane-1,2-diol (12): $[a]_{D}^{20} =$ +13 (c 1.3, CHCl₃). M.p. 50 °C (CH₂Cl₂). IR (film): $\tilde{v} = 3418$, 2956, 2929, 2887, 2857, 1471, 1362, 1252, 1045, 836 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.11$ (s, 6 H), 0.91 (s, 9 H), 1.10 (s, 3 H), 1.20 (s, 9 H), 1.26 (td, J = 3.2, 15.0 Hz, 1 H), 1.60 (br. s, 1 H), 1.75 (ddd, J = 4.0, 4.9, 13.3 Hz, 1 H), 1.91 (ddd, J = 2.0, 11.5, 13.3 Hz,1 H), 2.45 (td, J = 7.6, 15.0, Hz, 1 H), 2.70 (br. s, 1 H), 3.30 (dd, J = 2.3, 4.0 Hz, 1 H), 3.35 (d, J = 9.2 Hz, 1 H), 3.39 (s, 3 H), 3.65 (s, 1 H), 3.76 (m, 3 H), 3.77 (d, J = 9.2 Hz, 1 H), 4.20 (d, J = 5.2, 11.4 Hz, 1 H), 4.63 (d, J = 6.2 Hz, 1 H), 4.67 (d, J = 6.2 Hz, 1 H) ppm. ¹³C NMR (63 MHz, CDCl₃): $\delta = -4.9, -4.1, 18.0, 21.6, 25.8$ (3 C), 28.8 (3 C), 33.4, 39.2, 42.1, 55.5, 58.9, 67.2, 68.1, 70.8, 73.5, 74.1, 77.2, 97.0 ppm. ESIMS (MeOH): m/z (%) = 473.2 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for C₂₂H₄₆O₇NaSi 473.2911; found 473.2937. C₂₂H₄₆O₇Si (450.30)·0.25H₂O: calcd. C 57.28, H 10.16; found C 57.55, H 9.79.

5-*tert*-Butoxy-4-(2-hydroxyethyl)-2-methoxymethoxymethyl-4-methylcyclohexane-1,2,3-triol (13): $[a]_D^{20} = +20$ (c 1.0, MeOH). M.p. 50 °C (CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃): δ = 1.03 (s, 3 H), 1.13 (s, 9 H), 1.60 (td, J = 5.6, 14.6 Hz, 1 H), 1.76 (td, J = 4.3, 13.1 Hz, 1 H), 2.01 (ddd, J = 2.0, 12.0, 13.2 Hz, 1 H), 2.28 (ddd, J = 6.8, 8.2, 14.6 Hz, 1 H), 3.36 (s, 3 H), 3.54 (dd, J = 2.0, 4.3 Hz, 1 H), 3.54 (s, 1 H), 3.54 (d, J = 8.9 Hz, 1 H), 3.58 (d, J = 8.9 Hz, 1 H), 3.63 (m, 1 H), 3.71 (m, 1 H), 4.09 (dd, J = 4.7, 12.0 Hz, 1 H), 4.62 (s, 2 H) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 22.2, 29.1 (3 C), 33.7, 37.9, 43.5, 55.5, 59.6, 66.6, 69.8, 72.7, 74.4, 74.5, 77.4, 97.9 ppm. ESIMS (MeOH): m/z (%) = 359.2 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for C₁₆H₃₂O₇Na 359.2046; found 359.2044.

2-[5-tert-Butoxy-7-(tert-butyldimethylsilanyloxy)-7a-methoxymethoxymethyl-2,2,4-trimethyl-hexahydrobenzo[1,3]dioxol-4-yl]ethanol (14): To a stirred solution of 12 (190 mg, 0.42 mmol) in acetone (2 mL), dry Na₂SO₄ (1.0 g) and p-TsOH (20 mg) were added at 0 °C. The mixture was stirred at room temperature for 1 h 15 min (TLC monitoring) and then filtered through alumina. Purification by silica gel column chromatography (heptane/AcOEt, 7:1 to 3:1) afforded 173 mg (84%) of isopropylidene alcohol 14 and 8.3 mg (7%) of overprotection in the form of mixed acetal. Compound 14: $[a]_{D}^{20} = +25$ (c 0.6, CHCl₃). IR (film): $\tilde{v} = 3471$, 2955, 2930, 2857, 1255, 1081, 1041, 835 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.08$ (s, 3 H), 0.09 (s, 3 H), 0.92 (s, 9 H), 0.95 (s, 3 H), 1.19 (s, 9 H), 1.38 (s, 3 H), 1.50 (s, 3 H), 1.71 (ddd, J = 3.9, 8.6, 14.1 Hz, 1 H), 1.86 (m, 2 H), 1.95 (ddd, J = 5.0, 6.0, 14.1 Hz, 1 H), 2.60 (br. s, 1 H), 3.37 (s, 3 H), 3.48 (d, J = 10.5 Hz, 1 H), 3.58 (d, J = 10.5 Hz, 1 H), 3.77 (m, 2 H), 3.89 (dd, J = 3.9, 6.0 Hz, 1 H), 3.90 (dd, J =5.0, 8.6 Hz, 1 H), 4.13 (s, 1 H), 4.64 (d, J = 6.5 Hz, 1 H), 4.67 (d, J = 6.5, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta = -4.6, -4.2,$ 17.0, 18.7, 26.2, 26.5 (3 C), 26.9, 29.1 (3 C), 36.6, 39.0, 41.3, 55.5, 59.1, 68.5, 68.7, 70.1, 73.5, 81.3, 82.1, 96.8, 107.4 ppm. ESIMS (MeOH): m/z (%) = 513.3 (100) [M + Na]⁺. HRESIMS (MeOH):

calcd. for $C_{25}H_{50}O_7NaSi$ 513.3224; found 513.3208. $C_{25}H_{50}O_7Si$ (490.33)·0.2 C_7H_{16} : calcd. C 62.08, H 10.50; found C 62.24, H 10.28.

{6-tert-Butoxy-3a-methoxymethoxymethyl-7-[2-(1-methoxy-1-methylethoxy)-ethyl]-2,2,7-trimethyl-hexahydrobenzo[1,3]dioxol-4yloxy}tert-butyldimethylsilane: [a] $_{D}^{20}$ = +19 (c 1.0, CHCl $_{3}$).IR (film): $\tilde{v} = 2955, 2933, 2886, 2858, 1464, 1377, 1364, 1256, 1041, 837,$ 775 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.06$ (s, 3 H), 0.07 (s, 3 H), 0.91 (s, 9 H), 0.92 (s, 3 H), 1.18 (s, 9 H), 1.33 (s, 6 H), 1.35 (s, 3 H), 1.46 (s, 3 H), 1.67 (ddd, J = 4.0, 7.8, 14.0 Hz, 1 H), 1.73– 1.88 (m, 2 H), 1.94 (ddd, J = 5.2, 6.9, 14.0 Hz, 1 H), 3.19 (s, 3 H), 3.36 (s, 3 H), 3.46 (d, J = 10.5 Hz, 1 H), 3.47 (m, 1 H), 3.56 (d, J= 10.5 Hz, 1 H), 3.57 (m, 1 H), 3.83 (dd, J = 5.3, 7.8 Hz, 1 H), 3.90 (dd, J = 3.9, 6.9 Hz, 1 H), 4.00 (s, 1 H), 4.63 (d, J = 6.5 Hz,1 H), 4.65 (d, J = 6.5 Hz, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta = -4.5, -4.1, 17.3, 18.6, 24.4, 24.6, 26.1, 26.4 (3 C), 27.0, 29.1 (3 C)$ C), 36.8, 38.4, 38.6, 48.4, 55.5, 57.0, 68.5, 69.3, 70.3, 73.3, 81.1, 81.7, 96.9, 99.9, 107.2 ppm. ESIMS (MeOH): m/z (%) = 585.3 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for C₂₉H₅₈O₈NaSi 585.3799; found 585.3814. C₂₉H₅₈O₈Si (562.39)·0.3H₂O: calcd. C 63.00, H 10.68; found C 63.15, H 10.38.

Oxidation of 14 with Dess–Martin's Periodinane: To a solution of the above isopropylidene alcohol 14 (70 mg, 0.14 mmol) in dry dichloromethane (2 mL) and pyridine (0.12 mL, 1.4 mmol) was added periodinane (182 mg, 0.42 mmol). Stirring was continued at room temperature for 30 min. The reaction was then diluted with dichloromethane, quenched with a saturated aqueous solution of sodium hydrogencarbonate, washed with 5% aqueous Na₂S₂O₃ and then brine, and worked up as usual to give, after silica gel column chromatography (heptane/EtOAc, 10:1 to 4:1), 60 mg (86%) of 6 as an oil.

[5-tert-Butoxy-7-(tert-butyldimethylsilanyloxy)-7a-methoxymethoxymethyl-2,2,4-trimethylhexahydrobenzo[1,3]dioxol-4-yl]acetal**dehyde (6):** $[a]_D^{20} = +38$ (c 1.4, CHCl₃). IR (film): $\tilde{v} = 2934$, 2885, 2857, 1720, 1463, 1377, 1257, 1077, 1040, 836 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.08$ (s, 3 H), 0.09 (s, 3 H), 0.92 (s, 9 H), 1.07 (s, 3 H), 1.19 (s, 9 H), 1.34 (s, 3 H), 1.48 (s, 3 H), 1.71 (ddd, J = 3.9, 9.1, 14.1 Hz, 1 H), 1.95 (ddd, J = 5.1, 5.7, 14.1 Hz, 1 H), 2.49 (dd, J = 3.7, 15.4 Hz, 1 H), 2.62 (dd, J = 1.5, 15.4 Hz, 1 H),3.36 (s, 3 H), 3.49 (d, J = 10.4 Hz, 1 H), 3.56 (d, J = 10.4 Hz, 1 H), 3.91 (dd, J = 3.9, 5.7 Hz, 1 H), 4.01 (dd, J = 4.9, 9.1 Hz, 1 H), 4.21 (s, 1 H), 4.63 (d, J = 6.7 Hz, 1 H), 4.66 (d, J = 6.7 Hz, 1 H), 9.88 (dd, J = 1.5, 3.7 Hz, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta = -4.6, -4.2, 17.6, 18.6, 26.3, 26.5 (3 C), 27.0, 29.1 (3 C), 36.5,$ 40.0, 52.1, 55.6, 67.6, 68.9, 70.3, 73.8, 81.0, 82.1, 96.9, 107.8, 203.5 ppm. ESIMS (MeOH): m/z (%) = 511.3 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for C25H48O7NaSi 511.3067; found 511.3065. C₂₅H₄₈O₇Si (488.32)·0.35CH₂Cl₂: calcd. C 58.73, H 9.47; found C 58.52, H 9.45.

Selective Acetonide Formation from Tetraol 13: Proceeding as above, 13 (37 mg, 0.11 mmol) in acetone (1 mL), molecular sieves, dry Na₂SO₄ (500 mg), and *p*-TsOH (catalytic) at 0 °C to room temperature for 3 h afforded, after chromatography (heptane/EtOAc, 3:1 to EtOAc), single acetonide 15 (33 mg, 80%).

{6-tert-Butoxy-3a-methoxymethoxymethyl-7-[2-(1-methoxy-1-methylethoxy)ethyl]-2,2,7-trimethylhexahydrobenzo[1,3]dioxol-4-yloxy}tert-butyldimethylsilane (15): [a] $_{\rm D}^{20}$ = +22 (c 0.4, CHCl $_{\rm 3}$). IR (film): $\tilde{\rm v}$ = 3447, 2975, 2939, 2886, 1380, 1043 cm $^{-1}$. 1 H NMR (400 MHz, CDCl $_{\rm 3}$): δ = 0.97 (s, 3 H), 1.21 (s, 9 H), 1.44 (s, 3 H), 1.53 (s, 3 H), 1.75 (ddd, J = 4.0, 9.1, 14.4 Hz, 1 H), 1.88 (t, J = 6.2 Hz, 2 H), 2.06 (ddd, J = 4.8, 5.7, 14.4 Hz, 1 H), 2.29 (br. s, 1 H), 2.50 (br. s, 1 H), 3.38 (s, 3 H), 3.50 (d, J = 11.0 Hz, 1 H), 3.63 (d, J = 11.0 Hz, 1 H), 3.75 (dd, J = 3.5, 6.2 Hz, 1 H), 3.79 (m, 2

H), 3.92 (dd, J = 4.7, 8.9 Hz, 1 H), 4.23 (s, 1 H), 4.66 (d, J = 6.5 Hz, 1 H), 4.68 (d, J = 6.5 Hz, 1 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = 16.8$, 25.5, 26.5, 29.1 (3 C), 34.6, 38.9, 41.1, 55.5, 59.2, 67.8, 68.1, 70.1, 73.8, 81.1, 81.8, 96.8, 107.6 ppm. ESIMS (MeOH): m/z (%) = 399.2 (100) [M + Na]⁺. HR ESIMS: calcd. for C₁₉H₃₆O₇Na 399.2359; found 399.2354. C₁₉H₃₆O₇ (376.25) ·0.25H₂O: calcd. C 59.90, H 9.66; found C 59.81, H 9.55.

Cleavage of Lactone 11b by Basic Methanolysis – Preparation of Key C-Ring Precursors: K₂CO₃ (378 mg, 2.73 mmol) in MeOH/H₂O (9 mL, 8:1) was added onto 11b (445 mg, 0.91 mmol) at 0 °C under an argon atmosphere. The solution was stirred overnight at room temperature. Methanol was removed under reduced pressure. Water and HCl (1 N) were added to neutralize the mixture, and it was then extracted with AcOEt. The organic phase was washed with brine, dried with magnesium sulfate, and concentrated under reduced pressure. Purification by flash chromatography (heptane/AcOEt, 6:1 to AcOEt/MeOH, 9:1) afforded 303 mg (72%) of expected product 16a, 72 mg (18%) of fused-γ-lactone 5, 16 mg (4%) of recovered starting material 11b and its de-acetylated derivative 11d (15 mg, 4%).

6-tert-Butoxy-8-(tert-butyldimethylsilanyloxy)-9-hydroxy-1-methoxymethoxymethyl-5-methyl-cyclohexyl-2-oxabicyclo[3.3.1]nonan-3one (11d): $[a]_D^{20} = +19$ (c 0.7, CHCl₃). M.p. 70–71 °C (CH₂Cl₂). IR (film): $\tilde{v} = 3448$, 2956, 2929, 2856, 1734, 1718, 1465, 1253, 1115, 1064, 836 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): $\delta = 0.05$ (s, 3 H), 0.06 (s, 3 H), 0.87 (s, 9 H), 1.08 (s, 3 H), 1.18 (s, 9 H), 1.55 (ddd, J = 2.5, 11.6, 14.4 Hz, 1 H), 1.88 (ddd, J = 3.0, 4.9, 14.5 Hz, 1 H), 2.12 (d, J = 18.9 Hz, 1 H), 2.64 (d, J = 18.9 Hz, 1 H), 3.37 (t, J = 18.92.8 Hz, 1 H), 3.40 (s, 3 H), 3.82 (br. s, 1 H), 3.90 (s, 1 H), 3.92 (d, J = 10.4 Hz, 1 H), 3.99 (d, J = 10.4 Hz, 1 H), 4.03 (dd, J = 4.9, 11.6 Hz, 1 H), 4.66 (s, 2 H) ppm. 13 C NMR (75 MHz, CDCl₃): δ = -4.9, -4.2, 17.9, 21.9, 25.7 (3 C), 28.6 (3 C), 34.4, 37.7, 39.8, 55.8,67.5, 67.9, 69.2, 73.2, 74.1, 85.2, 97.3, 171.1 ppm. ESIMS (MeOH): m/z (%) = 469.2 (100) [M + Na]⁺. HRESIMS (MeOH): calcd. for C₂₂H₄₂O₇NaSi 469.2598; found 469.2591. C₂₂H₄₂O₇Si (446.27) ·0.5C₇H₁₆: calcd. C 61.66, H 10.15; found C 61.73, H 9.83.

[6-tert-Butoxy-4-(tert-butyldimethylsilanyloxy)-2,3-dihydroxy-3-methoxymethoxymethyl-1-methyl-cyclohexyllacetic Acid (16a): 1 H NMR (300 MHz, CDCl₃): δ = 0.10 (s, 6 H), 0.90 (s, 9 H), 1.18 (s, 9 H), 1.19 (s, 3 H), 1.26 (br. s, 2 H), 1.74 (ddd, J = 3.4, 5.5, 13.5 Hz, 1 H), 1.84 (ddd, J = 1.7, 11.0, 13.5 Hz, 1 H), 2.49 (d, J = 14.7 Hz, 1 H), 3.26 (d, J = 14.7 Hz, 1 H), 3.38 (s, 3 H), 3.48 (d, J = 9.6 Hz, 1 H), 3.67 (dd, J = 1.7, 3.4 Hz, 1 H), 3.71 (d, J = 9.4 Hz, 1 H), 3.78 (s, 1 H), 4.13 (dd, J = 5.5, 11.0 Hz, 1 H), 4.63 (d, J = 6.7 Hz, 1 H), 4.64 (d, J = 6.7 Hz, 1 H) ppm. 13 C NMR (75 MHz, CDCl₃): δ = -5.0, -4.1, 17.9, 21.4, 25.7 (3 C), 28.6, (3 C) 32.8, 40.3, 41.3, 55.6, 67.2, 70.8, 72.2, 73.6 (3 C), 73.9, 75.5, 97.1, 176.6 ppm. ESIMS (MeOH): mlz (%) = 487.3 (100) [M + Na]⁺.

4-*tert*-**Butoxy-6-**(*tert*-**butyldimethylsilanyloxy**)-7-**hydroxy-7-methoxymethoxymethyl-3a-methylhexahydrobenzofuran-2-one (5):** [a]²⁰₂₀ = +20 (c 0.4, CHCl₃). IR (film): \tilde{v} = 3489, 2954, 2929, 2856, 1784, 1039, 1020, 837 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ = 0.10 (s, 3 H), 0.12 (s, 3 H), 0.90 (s, 9 H), 1.15 (s, 3 H), 1.18 (s, 9 H), 1.75 (ddd, J = 5.2, 7.5, 14.1 Hz, 1 H), 1.81 (ddd, J = 3.1, 6.4, 14.1 Hz, 1 H), 2.09 (d, J = 16.6 Hz, 1 H), 2.79 (d, J = 16.6 Hz, 1 H), 2.81 (s, 1 H), 3.28 (d, J = 9.4 Hz, 1 H), 3.36 (s, 3 H), 3.54 (d, J = 9.4 Hz, 1 H), 3.90 (dd, J = 3.1, 7.5 Hz, 1 H), 4.18 (dd, J = 5.2, 6.4 Hz, 1 H), 4.20 (s, 1 H), 4.60 (d, J = 6.3 Hz, 1 H), 4.62 (d, J = 6.3 Hz, 1 H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = -5.0, -4.2, 17.9, 23.3, 25.8 (3 C), 28.6 (3 C), 34.7, 42.0, 43.3, 55.7, 65.6, 67.2, 69.0, 73.5, 73.9, 83.5, 96.8, 177.2 ppm. ESIMS (MeOH): m/z (%) = 469.2 (100) [M + Na]⁺. HRESIMS: calcd. for C₂₂H₄₂O₇NaSi 469.2598; found

469.2583. $C_{22}H_{42}O_7Si$ (446.27)·0.2 C_7H_{16} : calcd. C 60.22, H 9.76; found C 60.23, H 9.41.

Esterification of 16a: To a magnetically stirred solution of 16a (303 mg, 0.65 mmol) in diethyl ether (6 mL) and methanol (1 mL) was added TMSCHN₂ (1.6 mL, 3.27 mmol) at 0 °C. The mixture was stirred for 15 min at 0 °C, concentrated under reduced pressure, and purified by silica gel column chromatography (heptane/ AcOEt, 6:1 to 3:1) to afford 279 mg (90%) of expected product 16b together with γ -lactone 5 (16 mg, 5%).

Methyl [6-tert-Butoxy-4-(tert-butyldimethylsilanyloxy)-2,3-dihydroxy-3-methoxymethoxymethyl-1-methylcyclohexyl]acetate (16b): $[a]_{\rm D}^{20} = +28 \ (c \ 1.2, \ {\rm CHCl_3}). \ {\rm M.p. \ 61 \ ^{\circ}C} \ ({\rm CH_2Cl_2}). \ {\rm IR} \ ({\rm film}): \ \tilde{v} =$ 3490, 2954, 2930, 2857, 1734, 1464, 1253, 1148, 1061, 1041, 836, 777 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): $\delta = 0.11$ (s, 6 H), 0.90 (s, 9 H), 1.14 (s, 3 H), 1.19 (s, 9 H), 1.76 (td, J = 4.6, 13.5 Hz, 1 H), 1.89 (ddd, J = 1.9, 11.5, 13.5 Hz, 1 H), 2.45 (s, 1 H), 2.73 (d, J =14.9 Hz, 1 H), 2.87 (d, J = 14.9 Hz, 1 H), 2.99 (br. s, 1 H), 3.38 (s, 3 H), 3.39 (d, J = 9.4 Hz, 1 H), 3.66 (s, 3 H), 3.70 (d, J = 9.4 Hz, 1 H), 3.70 (s, 1 H), 3.89 (dd, J = 1.9, 3.9 Hz, 1 H), 4.21 (dd, J =5.1, 11.5 Hz, 1 H), 4.63 (d, J = 6.4 Hz, 1 H), 4.64 (d, J = 6.4 Hz, 1 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = -5.0, -4.1, 18.0, 21.8,$ 25.8 (3 C), 28.7 (3 C), 33.0, 38.1, 41.8, 51.3, 55.5, 67.0, 69.0, 70.9, 72.5, 73.6, 75.8, 97.0, 174.1 ppm. ESIMS (MeOH): m/z (%) = 501.2 (100) [M + Na]⁺. HRESIMS: calcd. for $C_{23}H_{46}O_8SiNa$ 501.2860; found 501.2856. C₂₃H₄₆O₈Si (478.29): calcd. C 57.71, H 9.69; found C 57.77, H 9.66.

DIBAL Reduction of 5: Diisobutylaluminium hydride (30% in toluene, 0.22 mL, 0.26 mmol) was added slowly to 5 (40 mg, 0.09 mmol) in toluene (1 mL) at -78 °C under an argon atmosphere. The reaction was stirred for 2 h while warming from -78 °C to -40 °C. The mixture was then quenched by the addition of saturated NH₄Cl (a few drops), extracted with ether, and washed with brine. The organic layer was concentrated under reduced pressure and SiO₂ flash chromatography of the residue (heptane/EtOAc, 5:1 to 3:1) gave 37.2 mg (93%) of lactol 17 as the major isomer; the minor lactol could not be characterized as it was isolated in an impure form.

4-*tert*-Butoxy-6-(*tert*-butyldimethylsilanyloxy)-7-methyl-7-methoxymethoxymethyl-3a-methyloctahydrobenzofuran-2,7-diol (17): IR (film): $\tilde{v} = 3445$, 2956, 2930, 2888, 2858, 1472, 1253, 1044, 836 cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ = 0.11 (s, 3 H), 0.12 (s, 3 H), 0.90 (s, 9 H), 1.07 (s, 3 H), 1.18 (s, 9 H), 1.63 (dd, J = 5.1, 12.9 Hz, 1 H), 1.70 (ddd, J = 5.0, 6.1, 13.7 Hz, 1 H), 1.87 (ddd, J= 2.7, 8.5, 13.7 Hz, 1 H), 1.87 (dd, J = 6.2, 12.9 Hz, 1 H), 3.16 (s, 1 H), 3.29 (d, J = 9.1 Hz, 1 H), 3.36 (s, 3 H), 3.59 (d, J = 9.1 Hz, 1 H), 3.75 (dd, J = 2.7, 6.2 Hz, 1 H), 3.39 (s, 1 H), 4.22 (dd, J =4.8, 8.4 Hz, 1 H), 4.58 (d, J = 11.5 Hz, 1 H), 4.60 (d, J = 6.2 Hz, 1 H), 4.62 (d, J = 6.2 Hz, 1 H), 5.48 (ddd, J = 5.1, 5.9, 11.5 Hz, 1 H) ppm. ¹³C NMR (75 MHz, CDCl₃): $\delta = -5.0$, -4.2, 18.0, 24.5, 25.8 (3 C), 28.7 (3 C), 34.3, 46.8, 47.7, 55.6, 66.3, 67.4, 69.5, 73.5, 73.6, 82.2, 96.9, 100.1 ppm. ESIMS (MeOH): m/z (%) = 471.2 (100) $[M + Na]^+$. HRESIMS: calcd. for $C_{22}H_{44}O_7NaSi$ 471.2754; found 471.2753. C₂₂H₄₄O₇Si (448.29)·0.15C₇H₁₆: calcd. C 59.71, H 10.09; found C 59.76, H 10.09.

Preparation of α-Alkoxyorganostannanes – C-10 Nucleophile Precursors: To a magnetically stirred solution of diisopropylamine (0.19 mL, 1.19 mmol) in dry THF (0.7 mL) under an argon atmosphere at 0 °C, nBuLi (1.6 м hexane solution, 0.81 mL, 1.35 mmol) was added dropwise. The solution was stirred for 10 min, nBu₃SnH (0.36 mL, 1.35 mmol) was added, and stirring was continued for 15 min at 0 °C. The reaction mixture was chilled to –78 °C before a solution of 6 (53 mg, 0.11 mmol) in dry THF (1.8 mL) was added

dropwise. The mixture was stirred at –78 °C for 30 min and then quenched with saturated NH₄Cl, diluted with Et₂O, and extracted. The combined organic layers were washed with brine, dried with magnesium sulfate, and concentrated under reduced pressure. *i*Pr₂NEt (0.29 mL, 1.66 mmol) was added, followed by MOMCl (0.08 mL, 1.11 mmol) 10 min later, at 0 °C on the crude residue thus obtained in dichloromethane (1 mL). The solution was stirred overnight at room temperature, quenched with H₂O (at 0 °C), and extracted with CH₂Cl₂. The organic phase was washed with water and saturated NaHCO₃ and worked up as usual to afford, after purification by flash chromatography (heptane/AcOEt, 1:0 to 5:1), a 75% combined yield of the desired α-alkoxyorgano stannanes, faster eluting isomer 18 (34 mg) and slower eluting isomer 19 (32 mg), in a 1:1 ratio.

{6-tert-Butoxy-3a-methoxymethoxymethyl-7-[2-methoxymethoxy-2-(tributylstannanyl)ethyl]-2,2,7-trimethylhexahydrobenzo[1,3]dioxol-4-yloxy}tert-butyldimethylsilane (18/19): Faster Eluting Isomer 18: $[a]_{D}^{20} = -13$ (c 1.0, CHCl₃). IR (film): $\tilde{v} = 2956$, 2928, 2856, 1463, 1377, 1252, 1146, 1042, 836 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ = 0.08 (s, 6 H), 0.86–0.94 (m, 9 H), 0.93 (s, 9 H), 0.95 (s, 3 H), 1.18 (s, 9 H), 1.25–1.40 (m, 6 H), 1.35 (s, 3 H), 1.48 (s, 3 H), 1.48–1.56 (m, 6 H), 1.60-1.69 (m, 6 H), 1.72 (ddd, J = 4.5, 5.8, 14.0 Hz, 1)H), 1.75 (dd, J = 3.1, 14.6 Hz, 1 H), 2.03 (ddd, J = 6.4, 8.5, 14.0 Hz, 1 H), 2.31 (dd, J = 12.0, 14.6 Hz, 1 H), 3.34 (s, 3 H), 3.37 (s, 3 H), 3.52 (d, J = 10.2 Hz, 1 H), 3.59 (d, J = 10.2 Hz, 1 H), 3.73 (t, J =5.8 Hz, 1 H), 4.05 (dd, J = 4.5, 8.9 Hz, 1 H), 4.31 (dd, J = 3.1, 11.3 Hz, 1 H), 4.39 (s, 1 H), 4.49 (d, J = 6.7 Hz, 1 H), 4.58 (d, J =6.7 Hz, 1 H), 4.62 (d, J = 6.5 Hz, 1 H), 4.64 (d, J = 6.5 Hz, 1 H) ppm. ¹³C NMR (100 MHz, CDCl₃): $\delta = -4.3, -4.1, 9.7, 13.6$ (3 C), 16.3, 17.5 (2 C), 18.5, 26.2, 26.4 (3 C), 26.9 (2 C), 27.0, 27.6, 27.9 (2 C), 29.2 (3 C), 29.3, 37.2, 40.8 (2 C), 55.4, 55.9, 67.6, 69.6, 69.7, 70.4, 73.3, 81.2, 82.5, 96.0, 96.9, 106.9 ppm. ESIMS (MeOH): m/z $(\%) = 847.4 (100) [M + Na]^+$. HRESIMS: calcd. for $C_{39}H_{80}O_8NaS$ iSn 847.4542; found 847.4497. C₃₉H₈₀O₈SiSn (824.46)·0.9C₇H₁₆: calcd. C 59.45, H 10.40; found C 59.44, H 10.09. Slower Eluting Isomer 19: $[a]_D^{20} = +16$ (c 0.3, CHCl₃).IR (film): $\tilde{v} = 2956$, 2927, 2856, 1463, 1377, 1257, 1147, 1041, 834 cm⁻¹, ¹H NMR (400 MHz, CDCl₃): $\delta = 0.08$ (s, 3 H), 0.09 (s, 3 H), 0.86–0.95 (m, 15 H), 0.93 (s, 9 H), 1.08 (s, 3 H), 1.21 (s, 9 H), 1.25–1.36 (m, 6 H), 1.37 (s, 3 H), 1.46 (s, 3 H), 1.47-1.56 (m, 6 H), 1.71 (ddd, J = 3.7, 8.7, 14.1 Hz, 1H), 1.88 (d, J = 14.3 Hz, 1 H), 1.91 (ddd, J = 5.0, 5.9, 14.1 Hz, 1 H), 2.26 (dd, J = 11.3, 14.3 Hz, 1 H), 3.37 (s, 3 H), 3.38 (s, 3 H), 3.49 (d, J = 10.5 Hz, 1 H), 3.60 (d, J = 10.5 Hz, 1 H), 3.81 (dd, J= 5.0, 8.8 Hz, 1 H), 3.91 (dd, J = 3.7, 5.9 Hz, 1 H), 4.20 (s, 1 H),4.44 (d, J = 11.0 Hz, 1 H), 4.54 (d, J = 6.5 Hz, 1 H), 4.56 (d, J =6.5 Hz, 1 H), 4.66 (s, 2 H) ppm. $^{13}\mathrm{C}$ NMR (100 MHz, CDCl3): δ = -4.5, -4.1, 9.5 (3 C), 13.7 (3 C), 17.4, 18.7, 26.2, 26.6 (3 C), 27.0,27.6 (3 C), 29.2 (3 C), 29.4, 29.7 (2 C), 36.7, 40.7, 43.6, 55.4, 56.2, 69.0, 69.4, 70.5, 71.3, 73.3, 81.1, 81.2, 96.9, 97.0, 107.1 ppm. ES-IMS (MeOH): m/z (%) = 847.4 (100) [M + Na]⁺. HRESIMS: calcd. for C₃₉H₈₀O₈NaSiSn 847.4542; found 847.4564. C₃₉H₈₀O₈SiSn (824.46)·1.1C₇H₁₆: calcd. C 59.97, H 10.52; found C 60.16, H 10.04.

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